

Synthesis of Chemically Stable PGI₂ Analogs II
-Synthesis of Halogen Substituted PGI₂ Analogs-†‡

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Abstract--Syntheses of several stable PGI₂ analogs substituted by halogen atoms(s) at C-5 or(and) C-7 are described. Reaction of protected PGI₂ methyl ester (1a) with N-chlorosuccinimide gave 5-chloro- Δ^6 -PGI₁ derivative (2a), which was transformed into 5-chloro- and 5,7-dichloro-PGI₂ [(7) and (5)] by subsequent isomerization or chlorination. Similarly, reaction of 1b with N-bromosuccinimide gave 5-bromo- Δ^6 -PGI₁ derivative (3b), which was further transformed into 7-fluoro-PGI₂ (16) by silver fluoride treatment. These halogenated PGI₂ analogs were found to be much more stable than PGI₂.

Prostacyclin (PGI₂) is a useful therapeutic agent in cardiovascular field because of its powerful vasoactive properties.¹ However its chemical instability against hydrolysis² is disadvantageous for its practical use, and chemically stable PGI₂ analogs have been proposed as more practical and useful agents.³

The instability of PGI₂ molecule is due to the unstable enol ether linkage in the molecule. Since this enol ether group constitutes a part of the strained bicyclo(3.3.0)octane ring system, it is hydrolyzed much easier than an ordinary enol ether group.⁴ In fact, PGI₂ is easily hydrolyzed to result in the formation of biologically less active 6-oxo-PGF_{1 α} .² PGI₂ analogs with an electron-withdrawing group on or adjacent to the enol ether double bond were considered to be resistant against the hydrolysis since the electron-withdrawing group would reduce the electron density of the enol ether double bond to prevent it from the attack of the hydronium ion.⁵ These analogs should be chemically more stable than naturally occurring PGI₂. Along with this strategy, 5-phenylthio-PGI₂⁶ was synthesized by the electrophilic reaction of benzenesulfonyl chloride to the enol ether linkage of PGI₂, and 7-hydroxy-6(b),⁷ and 7-fluoro-PGI₂^{6(b),7} were also synthesized in our laboratory. Other stable PGI₂ analogs such as 7-oxo-PGI₂,⁸ 4-oxo-PGI₂,⁹ 5-oxo- Δ^6 -PGI₁,¹⁰ 5-cyano-PGI₂,¹¹ 5-carboxy-PGI₂,¹² 5-formyl-PGI₂,¹³

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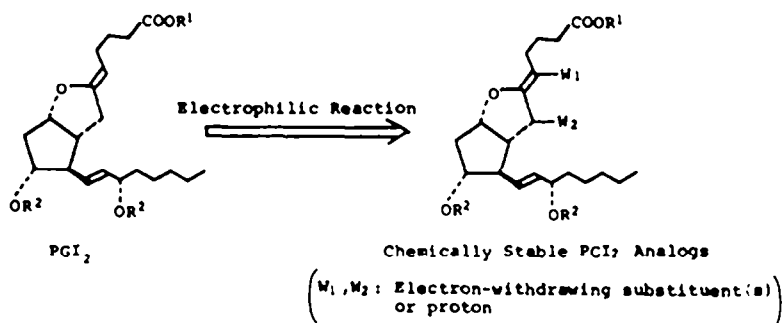
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10,10-difluoro-PGI₂¹⁴ etc.³ are also included in this category. In this report, electrophilic halogenation to the PGI₂-enol ether was examined, and syntheses of 5-chloro-, 5,7-dichloro-, and 7-fluoro-PGI₂ derivatives are described.

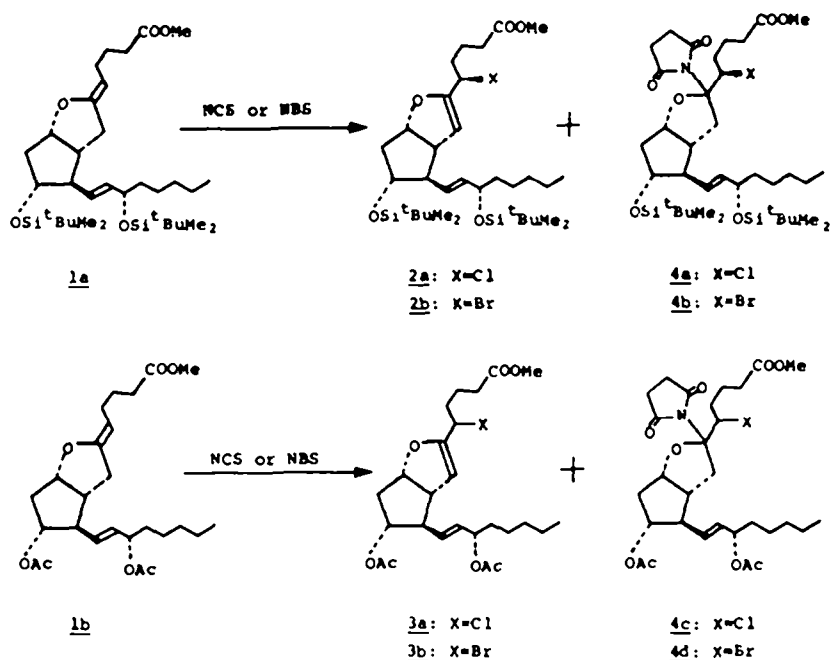
(I) Electrophilic halogenation of PGI₂ methyl ester

Previously we reported the synthesis of stable 5-phenylthio-PGI₂,⁶ where the electron-withdrawing phenylthio group on the enol ether double bond participated in the stabilization of the molecule against hydrolysis. However, the phenylthio substitution resulted in the decreased biological activity. As it was considered that the decreased activity might be due to the bulkiness of the phenylthio group, halogen atoms smaller than phenylthio function were chosen as electron-withdrawing groups for the chemical stabilization of PGI₂.



Scheme I

In order to introduce halogen atom into the PGI₂ skeleton, the electrophilic reaction of *N*-halosuccinimide with PGI₂-enol ether linkage was examined. Electrophilic chlorination of PGI₂ *bis*(*t*-butyldimethylsilyl ether) methyl ester (**1a**) with *N*-chlorosuccinimide (1.05 equiv.) was carried out in carbon tetrachloride, and (5*R*)-5-chloro-Δ⁶-PGI₁ *bis*(*t*-butyldimethylsilyl ether) methyl ester (**2a**) was obtained in 60% yield accompanied with the formation of succinimido-adduct **4a** (20%). Similarly, the reaction of **1a** with *N*-bromosuccinimide gave (5*R*)-5-bromo-Δ⁶-PGI₁ *bis*(*t*-butyldimethylsilyl ether) methyl ester (**2b**) in 57% yield accompanied also with succinimido-adduct **4b** (20%). Both allylic halides **2a** and **2b** were found to be homogeneous by the examinations of their ¹³C-NMR spectra, and their stereochemistry at C-5 was tentatively assigned as 5*R* (*vide infra*). On the other hand, the similar electrophilic halogenations of PGI₂ methyl ester diacetate (**1b**) with *N*-chlorosuccinimide or *N*-bromosuccinimide gave a diastereomeric mixture¹⁵ of 5-chloro-Δ⁶-PGI₁ diacetate methyl ester (**3a**) or 5-bromo-Δ⁶-PGI₁ diacetate methyl ester (**3b**) in 58% or 60% yield, respectively. Succinimido-adduct **4c** or **4d** was also obtained in these reactions in 20% or 13% yield, respectively. ¹³C-NMR spectra of the products **3a** and **3b** exhibited several pairs of peaks corresponding to the several carbons with the peak ratio of *ca.* 5:3 and *ca.* 7:3, respectively. These observations suggested that acetyl-protected allylic halides **3a** or **3b** were epimeric mixtures of (5*R*)- and (5*S*)-isomers in the respective ratios.



Scheme II

The exclusive formation of 5R-isomer of silyl-protected allylic halide **2a** or **2b** from silyl-protected PGI₂ methyl ester **1a** could be accounted for the bulkiness of the *t*-butyldimethylsilyl ether substituent at C-11. From the molecular model study, the C-11 substituent was found to be close to the enol ether double bond as depicted in Fig. 1.¹⁶ The *t*-butyldimethylsilyl group is bulky enough to hinder

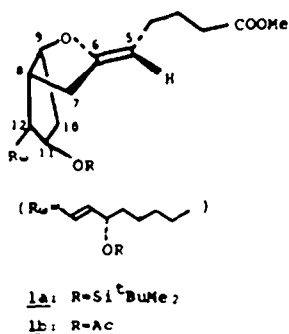
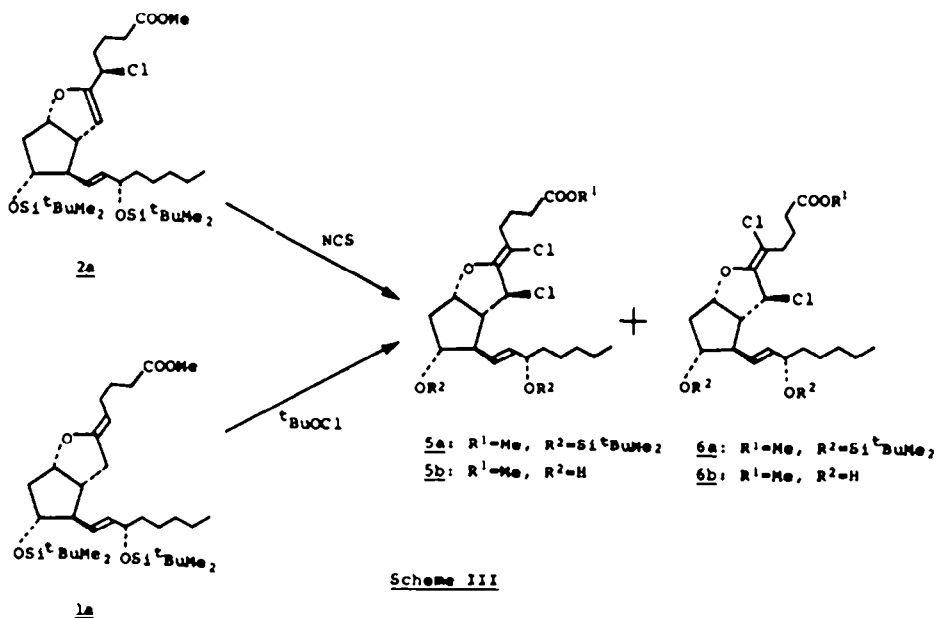


Fig. I

the lower side of the enol ether group from the attack of the halogenating agent, and consequently the halogenating agent could only approach from the less hindered upper side of the enol ether linkage to result in the exclusive formation of (5R)-isomers 2a or 2b. On the other hand, in the case of less hindered acetyl-protected PGI₂ 1b, the halogenating agent could approach from the both sides to give the epimeric mixture of (5R)- and (5S)-allylic halides. The stereochemistry at C-5 of silyl-protected succinimido-adducts 4a and 4b were tentatively assigned to 5R, whereas that of acetyl-protected adducts 4c and 4d were assumed to be mixtures of 5R and 5S.

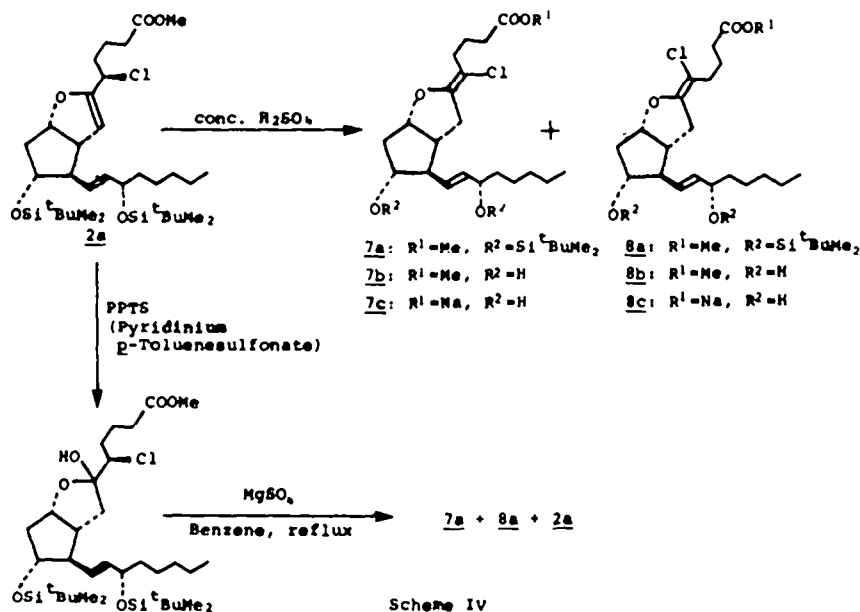
Further chlorination of silyl-protected (5R)-allylic chloride 2a was feasible with N-chlorosuccinimide. When allylic chloride 2a was treated with N-chlorosuccinimide in carbon tetrachloride-methylene chloride, geometric isomers, (5E,7S)- and (5Z,7S)-5,7-dichloro-PGI₂ bis(*t*-butyldimethylsilyl ether) methyl esters (5a) and (6a), were obtained in 17% and 20% yield, respectively. These dichlorides 5a and 6a were also obtained directly from silyl-protected PGI₂ methyl ester 1a by treatment with *t*-butyl hypochlorite¹⁷ (8 equiv.) in methylene chloride in 17% and 5% yield, respectively. The 7S-stereochemistry of 5a and 6a was determined from the ¹H-NMR study where the 7 α -methine protons of the products coupled with their 8 β -methine protons by less than one Hz suggesting the 7S-configuration of 5a and 6a.⁷ The Δ^5 -olefin geometry of 5a and 6a was also tentatively assigned from their ¹H-NMR studies as follows. The 7 α -methine proton of more polar dichloride 5a appeared in a field lower by 0.10 ppm than that of less polar 6a. By analogy with the case of 5-chloro-PGI₂ derivatives 7b and 8b (vide *infra*) where 7 α -methine proton of 5E-isomer 7b appeared in a field lower by 0.11 ppm than that of 5Z-isomer 8b, the more polar dichloride 5a and the less polar 6a were tentatively assigned to 5E and 5Z, respectively.

Removal of the silyl-protecting groups of 5a and 6a with hydrogen fluoride-pyridine in acetonitrile afforded (5E,7S)- and (5Z,7S)-5,7-dichloro-PGI₂ methyl ester (5b) and (6b) in 80% and 76% yield, respectively.



(II) Isomerization of 5-chloro- Δ^6 -PGI₁ into 5-chloro-PGI₂

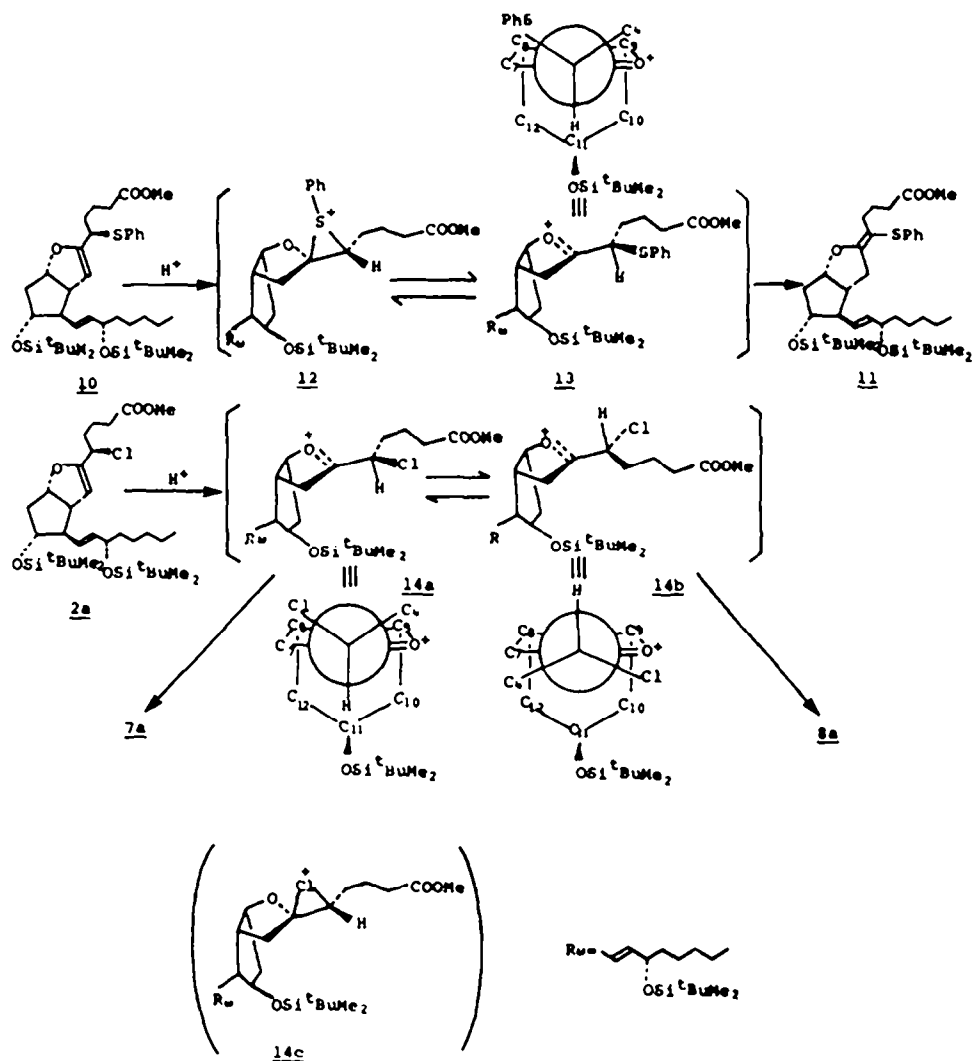
Isomerization of the endo-double bond (Δ^6) of allylic chloride 2a into the exo-double bond (Δ^5) is an important transformation to get a stable PGI₂ analog such as 5-chloro-PGI₂ (7). Previously, acid catalyzed stereospecific isomerization of silyl-protected 5-phenylthio- Δ^6 -PGI₁ methyl ester 10 into (5E)-5-phenylthio-PGI₂ derivative 11 was reported.⁶ Similar treatment of silyl-protected (5R)-5-chloro- Δ^6 -PGI₁ 2a with a trace amount of concentrated sulfuric acid gave a mixture of 5E- and 5Z-vinylic chlorides 7a and 8a only in low yield (both in 10% yields) after



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chromatographic separation. The difference in the stereochemical results would be attributed to the different intermediate formations as follows. In the acid catalyzed isomerization of 5-phenylthio derivative 10, there was a proposal of an exclusive formation of episulfonium ion intermediate 12 which could lead only to (5E)-5-phenylthio derivative via oxonium ion intermediate 13.^{6(b)} The type 14b conformer of 13 would be excluded probably due to the restricted bond rotation between C-5 and C-6 by the contribution of episulfonium ion 12 or by the bulkiness of the phenylthio group. On the other hand, in the case of the isomerization of 5-chloro derivative 2a, two possible conformational isomers of oxonium ion intermediates 14a and 14b were considered to be more plausible than three membered ring intermediate such as epichloronium intermediate 14c. These two intermediates 14a and 14b would lead to the formation of 7a and 8a, respectively.

The alternative formation of 5-chloro-PGI₂ derivative 7a was achieved by stepwise transformation of 2a via hemiacetal 9. When allylic chloride 2a was treated with a catalytic amount of pyridinium p-toluenesulfonate in moistured benzene, 5-chloro-6-hydroxy-PGI₁ derivative 9 was obtained as an epimeric mixture at C-6. Subsequent dehydration of the crude hemiacetal 9 with excess anhydrous magnesium sulfate in refluxing benzene resulted in the formation of 7a and 8a along with the starting allylic chloride 2a in 20%, 28% and 18% yields, respectively. Removal of the silyl protecting groups of 7a and 8a with tetrabutylammonium fluoride in tetrahydrofuran afforded (5E)-5-chloro-PGI₂ methyl ester (7b) and



Scheme V

(5E)-5-chloro-PGI₂ methyl ester (**8b**) in 95% and 80% yields, respectively. Hydrolysis of **8b** and **9b** with sodium hydroxide in ethanol-water gave sodium salt solution of (5E)-5-chloro-PGI₂ **7c** and (5Z)-5-chloro-PGI₂ **8c**, which were used directly for pharmacological tests.

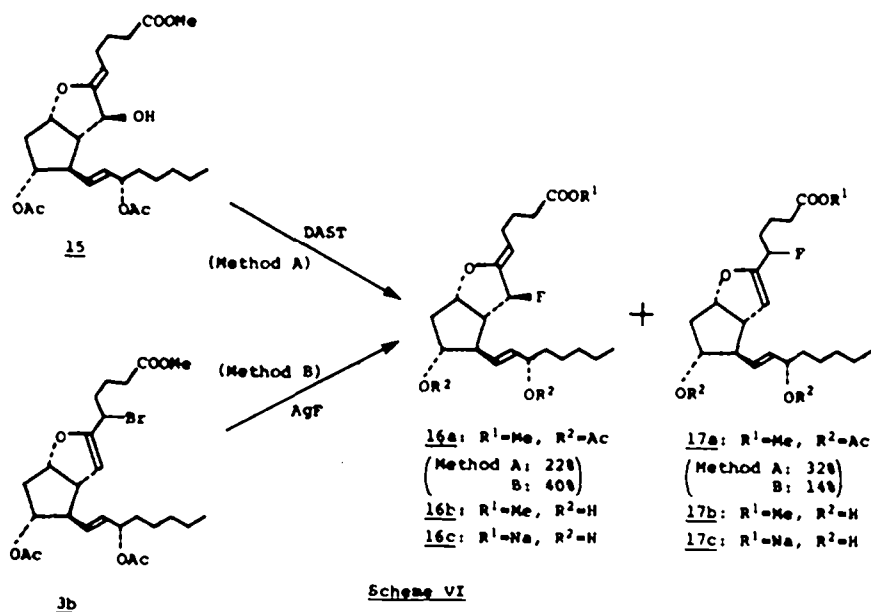
The Δ^5 -olefin geometries of the products **7b** and **8b** were determined on the basis of their $^1\text{H-NMR}$ (400 MHz) spectra with the aid of decoupling studies. The C-4 methylene protons of less polar component **7b** appeared in a field lower by 0.21 ppm (1H) and 0.11 ppm (1H) than those of more polar component **8b**. Since allylic methylene protons *cis* to oxygen atom are known to appear lower field than those of *trans*,¹⁸ the Δ^5 -olefin geometry of less polar isomer **7b** was assigned to 5E, and more polar **8b**, to 5Z.

(III) Fluorination of 5-halo- Δ^6 -PGI₁ into 7-fluoro-PGI₂

Fluorine atom substituted in the vicinity of the enol ether group of PGI₂ molecule should stabilize the enol ether linkage effectively because of the strong electron-withdrawing character.¹⁹ Moreover, it is known that the substitution by fluorine atom of biologically active substances often leads to their enhanced biological activities.²⁰ In these respects, we have already reported the synthesis of chemically stable 7-fluoro-PGI₂ (16c),⁷ which also retained its biological activity. However the previous method involving the fluorination of 7-hydroxy-PGI₂ derivative 15 with diethylaminosulfur trifluoride (DAST) is not satisfactory because of the low yield.^{6(b)} The more convenient synthesis of 7-fluoro-PGI₂ (16) is the use of the present allylic bromide 3b.

Treatment of acetyl protected 5-bromo- Δ^6 -PGI₁ methyl ester (3b) with silver fluoride (10 equiv.) in acetonitrile gave (5*Z*,7*S*)-7-fluoro-PGI₂ methyl ester diacetate (16a) (40%) and 5-fluoro- Δ^6 -PGI₁ methyl ester 11,15-diacetate (17a) (20%) after chromatographic separation. Similar reactions of allylic chloride 3a with silver fluoride or of allylic bromide 3b with cesium fluoride (in DMF, r.t. \sim 160°C) gave poor results to obtain the expected allylic fluoride 16a. The Δ^5 -double bond geometry and the C-7 stereochemistry of 7-fluoro-PGI₂ derivative 16a thus obtained was determined as (5*Z*,7*S*) by comparison with the authentic sample synthesized previously.^{6(b)} The isolated 5-fluoro- Δ^6 -PGI₁ derivative 17a was an inseparable mixture of epimers at C-5.

Removal of the acetyl groups of 16a by treatment with sodium methoxide in absolute methanol afforded (7*S*)-7-fluoro PGI₂ methyl ester (16b) in 82% yield, and hydrolysis of 16b with sodium hydroxide in ethanol-water gave sodium salt solution of (7*S*)-7-fluoro-PGI₂ (16c). The present new method is a convenient way to synthesize 7-fluoro-PGI₂ (16c) from PGI₂ methyl ester diacetate (1b).



Halogenated PGI₂ analogs thus obtained were found much more stable than PGI₂. The chemical half life of 5-chloro- and 5,7-dichloro-PGI₂ methyl esters **7b**, **8b**, **5b** and **6b** in pH 2.4 buffer solution was 4h, 6h, 12h and 2.5 h, respectively, while that of PGI₂ methyl ester was less than one minute. Moreover 7-fluoro-PGI₂ **16c** had a half-life more than one month in pH 7.4 buffer solution,^{6(b)} while that of PGI₂ in pH 7.46 has been reported to be only 15 minutes. 7-Fluoro-PGI₂ **16c** and **(5E)**-5-chloro-PGI₂ **7c** showed inhibitory activities on platelet aggregation (Rabbit, IC₅₀: 0.05 and 0.14 µg/ml, respectively), while **(5E)**-5-chloro-PGI₂ **8c** and 5,7-dichloro-PGI₂ methyl esters **5b** and **6b** were less active (>10 µg/ml).

Experimental

IR spectra were recorded on a JASCO A102 spectrometer ¹H- and ¹³C-NMR spectra were obtained on a JEOL JNM-GX400 (400 MHz), JEOL JNM-PS-100 (100 MHz) or a Varian EM360A (60 MHz) spectrometer. Chemical shifts are reported as parts per million (ppm) relative to internal tetramethylsilane. Mass spectra were taken at 70 or 20 eV on a HITACHI M-80B or a LKB-9000 mass spectrometer. Optical rotations were measured on a Union Giken PM-101 automatic polarimeter. For high pressure liquid chromatography (HPLC) analysis, a Shimadzu Model LC-3A liquid chromatography equipped with a Shimadzu SPD-2A UV detector, and a YMC-Pack A-312 ODS (5 µm) column (15 cm × 6 mm I.D.) were employed. Thin layer chromatography (TLC) was performed using Merck silica gel (Kieselgel 60 F₂₅₄) analytical or preparative plates. All reactions were carried out under argon or nitrogen. Solvents for reactions were purified if necessary before use by distillation from suitable drying agents. Solvents for extraction and chromatography were GR grades.

Chlorination of PGI₂ bis(t-butyldimethylsilyl ether) methyl ester (1a) with N-chlorosuccinimide (NCS) — (5R)-5-Chloro-δ⁶-PGI₂ bis(t-butyldimethylsilyl ether) methyl ester (2a) and 5-chloro-6-succinimido-PGI₂ bis(t-butyldimethylsilyl ether) methyl ester (4a)

N-Chlorosuccinimide (17 mg, 0.13 mmol) was added at room temperature to a stirred solution of **1a** (71 mg, 0.12 mmol) in carbontetrachloride (4 ml) in the presence of solid potassium carbonate (25 mg). After stirring the mixture at room temperature for 1 hr, water (10 mL) was added and the mixture was extracted with ether (2 × 20 mL). The combined extracts were washed with brine, dried over MgSO₄·K₂CO₃, and concentrated *in vacuo*. The residue was chromatographed on Florisil eluting with n-hexane-ethyl acetate (99.5 : 0.5) containing 0.1% of triethylamine to give allylic chloride **2a** (45 mg, 60%) as an oil; [α]_D²³ + 31° (c, 0.17, CHCl₃); IR (film) 1742, 1658, 1255, 1122, 1090, 1002, 970, 838, 775 cm⁻¹; ¹H-NMR (CDCl₃) δ 2.85-3.2(1H, br), 3.66(3H, s), 3.7-4.2(2H, m), 4.38(1H, t, J=7 Hz), 4.7-5.0(1H, br), 5.00(1H, d, J=3 Hz), 5.52 (2H, m); ¹³C-NMR (CDCl₃) δ 33.3(C₂), 22.0(C₃), 34.9(C₄), 55.4(C₅), 154.8(C₆), 101.6(C₇), 50.5(C₈), 83.3(C₉), 42.7(C₁₀), 76.1(C₁₁), 57.9(C₁₂); MS m/e 630, 628(M⁺), 592, 573, 571, 535. Calc. for C₃₃H₆₁NO₅Si₂ ³⁵Cl 628.3743, Found 628.3780.

Further elution with n-hexane-ethyl acetate (8 : 2) containing 0.1% of triethylamine gave succinimido-adduct **4a** (17.5 mg, 20%) as an oil; IR (CHCl₃) 1730, 1708, 1338, 1255, 972, 838 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.03(12H, s), 0.87(9H, s), 0.88(9H, s), 2.66(4H, s), 3.66(3H, s), 3.7-4.2(2H, m), 4.45-4.9(2H, m), 5.46(2H, m); MS m/e 672, 670(M⁺-tBu), 630, 628, 592, 573, 571, 535. Calc. for C₃₃H₅₇NO₇Si₂ ³⁵Cl 670.3358, Found 670.3322.

Chlorination of PGI₂ diacetate methyl ester (1b) with N-chlorosuccinimide (NCS) — 5-Chloro-δ⁶-PGI₂ diacetate methyl ester (3a) and 5-chloro-6-succinimido-PGI₂ diacetate methyl ester (4c)

Chlorination of **1b** (125 mg, 0.28 mmol) with NCS (39 mg, 0.29 mmol) was carried out according to the same method described above. The crude product was purified by Florisil column chromatography to isolate two compounds; **3a** (78 mg, 58%) (eluting solvent : n-hexane-ethyl acetate, 87.5 : 12.5, 0.1% triethylamine) and **4c** (32 mg, 20%) (eluting solvent : ethyl acetate, 0.1% triethylamine); **3a** IR (film) 1735, 1660, 1370, 1240, 1060, 1018, 968 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.88(3H, t, J=5 Hz), 1.98(3H, s), 2.00(3H, s), 2.95-3.4(1H, m), 3.69(3H, s), 4.39(1H, bt, J=6 Hz), 4.7-5.35(4H, s), 5.53(2H, m); ¹³C-NMR (CDCl₃) δ 33.2(C₂), 22.1 and 22.0 (C₃), 34.3(C₄), 54.3 and 54.5 (C₅), 155.0 and 155.4(C₆), 101.3 and 101.0(C₇), 51.5 (C₈), 84.9 and 84.6(C₉), 38.7(C₁₀), 78.03 and 77.79(C₁₁), 54.9(C₁₂); MS m/e 486, 484(M⁺), 448, 426, 424, 388, 366, 364, 329, 328. Calc. for C₂₅H₃₇O₇³⁵Cl 484.2226, Found 484.2192. **4c**; ¹H-NMR (CDCl₃) δ 0.88(3H, t, J=5 Hz), 2.00(6H, s), 2.65(4H, s), 2.65(4H, s), 3.67(3H, s), 4.4-5.3(4H, m), 5.50(2H, m).

Bromination of PGI₂ bis(t-butyldimethylsilyl ether) methyl ester (1a) with N-bromosuccinimide (NBS) — (5R)-5-bromo-δ⁶-PGI₂ bis(t-butyldimethylsilyl ether) methyl ester (2b) and 5-bromo-6-succinimide-PGI₂ bis(t-butyldimethylsilyl ether) methyl ester (4b)

Bromination of **1a** (60 mg, 0.10 mmol) with NBS (18.7 mg, 0.105 mmol) was carried out by the similar method described above. The crude product was purified by Florisil column chromatography (n-hexane-ethyl acetate, 99.5 : 0.5, 0.1% triethylamine) to give allylic bromide **2b** (38 mg, 57%) as an oil; $[\alpha]_D^{25} + 59^\circ$ (c, 0.32, CHCl₃); IR (film) 1742, 1254, 1122, 968, 835, 775 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.04(12H, s), 0.86(9H, s), 0.89(9H, s), 2.92(1H, t, J=8 Hz), 3.68(3H, s), 3.65-3.9(1H, m), 3.9-4.15(1H, m), 4.47(1H, t, J=7 Hz), 4.7-5.0(1H, m), 5.01(1H, d, J=2 Hz), 5.47(2H, m); ¹³C-NMR (CDCl₃) δ 33.2(C₂), 23.3(C₃), 35.5(C₄), 46.1(C₅), 155.3(C₆), 101.6(C₇), 50.6(C₈), 83.1(C₉), 42.6(C₁₀), 76.0(C₁₁), 57.8(C₁₂); MS m/e 674, 672(M⁺), 617, 615, 592, 537, 535, 460. Calc. for C₃₃H₆₁O₅Si₂⁷⁹Br 672.3238, Found 672.3181.

Further elution with n-hexane-ethyl acetate (8 : 2) containing 0.1% of triethylamine gave an oily succinimido-adduct **4b** (15.5 mg, 20%) IR (film) 1740, 1712, 1258, 975, 838, 778 cm⁻¹; ¹H-NMR (CHCl₃) δ 0.03(12H, s), 0.84(21H, bs), 2.63(4H, s), 3.65(3H, s), 3.6-4.2(2H, m), 4.3-5.0(2H, m), 5.48(2H, m); MS m/e 716, 714 (M⁺-tBu), 674, 672, 634, 617, 615, 553, 535. Calc. for C₃₃H₅₇NO₇Si₂⁷⁹Br (M⁺-tBu) 714.2854, Found 714.2939.

Bromination of PGI₂ diacetate methyl ester (**1b**) with N-bromosuccinimide (NBS)—5-bromo-6^b-PGI₂ diacetate methyl ester (**3b**) and 5-bromo-6-succinimido-PGI₂ diacetate methyl ester (**4d**)

Bromination of **1b** (372 mg, 0.83 mmol) with NBS (155 mg, 0.87 mmol) was carried out by following the same method described above. The crude product was purified by Florisil column chromatography (n-hexane-ethyl acetate, 85 : 15, 0.1% triethylamine) to give allylic bromide **3b** (262 mg, 60%) as an oil; $[\alpha]_D^{25} + 39.5^\circ$ (c, 0.51, CHCl₃); IR (film) 1740, 1655, 1372, 1240, 1060, 1018, 968 cm⁻¹; ¹H-NMR (CDCl₃) 0.88(3H, bt, J=5 Hz), 1.99(3H, s), 2.00(3H, s), 2.9-3.3(1H, m), 3.67(3H, s), 4.41(1H, bt, J=7 Hz), 4.6-5.3(4H, m), 5.48(2H, m); ¹³C-NMR (CDCl₃) δ 33.1(C₂), 23.2(C₃), 35.4 and 35.6(C₄), 45.5(C₅), 155.8 and 155.4(C₆), 101.0 and 101.2(C₇), 51.6(C₈), 84.5 and 84.7(C₉), 38.8(C₁₀), 77.6 and 77.9(C₁₁), 54.4 and 54.2(C₁₂); MS m/e 530, 528 (M⁺), 470, 468, 449, 448, 389, 388, 329. Calc. for C₂₅H₃₇O₇⁷⁹Br 528.1721, Found 528.1646.

Further elution with ethyl acetate containing 0.1% of triethylamine gave succinimido-adduct **4d** (66 mg, 13%) as an oil; IR (film) 1740, 1710, 1438, 1372, 1330, 1250, 1010, 975, 918, 732 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.87(3H, t, J=5 Hz), 2.00(6H, s), 2.68(4H, s), 3.67(3H, s), 4.4-5.3(4H, m), 5.51(2H, m); MS m/e 530, 528 (M⁺-succinimide), 448.

Further chlorination of (5R)-5-chloro-6^b-PGI₂ bis(t-butyl dimethylsilyl ether) methyl ester (**2a**) with NCS—(5E,7S)- and (5Z,7S)-5,7-dichloro-PGI₂ bis(t-butyl dimethylsilyl ether) methyl esters (**5a**) and (**6a**)

N-Chlorosuccinimide (36 mg, 0.27 mmol) was added at room temperature to a stirred solution of **2a** (57 mg, 0.091 mmol) in carbon tetrachloride-dichloromethane (0.6 ml each) in the presence of solid potassium carbonate (38 mg). After stirring at room temperature (1.5 hr), the reaction mixture was filtered through celite which was washed with ethyl acetate (15 mL). The filtrate was washed with water (10 mL) and the water layer was extracted with ethyl acetate (10 mL) again. The combined organic layer was washed with brine twice, dried over MgSO₄-K₂CO₃, and concentrated *in vacuo*. The residue (61 mg) was separated by preparative TLC developing with 10% ethyl acetate-n-hexane containing 0.1% of triethylamine to give more polar dichloride **5a** (9.8 mg, 17%) and less polar dichloride **6a** (11.3 mg, 20%); **5a** $[\alpha]_D^{23} + 64^\circ$ (c, 0.49, CHCl₃); IR (film) 1750, 1682, 1255, 1122, 1180, 970, 835, 775 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.7-1.0(21H), 3.67(3H, s), 3.85(1H, q, J=8 Hz), 4.05-4.3(1H, br), 4.79(1H, s), 4.8-5.1(1H, br), 5.53(2H, m); ¹³C-NMR (CDCl₃) δ 32.7(C₂), 22.3(C₃), 31.6(C₄), 111.4(C₅), 152.5(C₆), 58.5(C₇), 56.1(C₈), 83.9(C₉), 41.2(C₁₀), 77.7(C₁₁), 52.6(C₁₂); MS m/e 607, 605 (M-57), 571, 569. Calc. for C₂₉H₅₁O₂Si₂³⁵Cl₂ (M-57) 605.2651, Found 605.2642. **6a** $[\alpha]_D^{23} + 79^\circ$ (c, 0.57, CHCl₃); IR (film) 1742, 1682, 1252, 1124, 970, 835, 775 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.7-1.0(21H), 3.67(3H, s), 3.86(1H, q, J=7 Hz), 3.9-4.2(1H, br), 4.69(1H, s), 4.85-5.1(1H, br), 5.53(2H, m); ¹³C-NMR (CDCl₃) δ 32.7(C₂), 22.6(C₃), 33.3(C₄), 108.7(C₅), 151.0(C₆), 57.6(C₇), 56.9(C₈), 83.2(C₉), 41.0(C₁₀), 77.8(C₁₁), 52.7(C₁₂); MS m/e 607, 605 (M-57), 571, 569. Calc. for C₂₉H₅₁O₂Si₂³⁵Cl₂ (M-57) 605.2651, Found 605.2622.

Dichlorination of **1a** with t-butylhypochlorite—**5a** and **6a**

t-Butylhypochlorite (42 μL, 0.37 mmol) was added at -40°C to a solution of **1a** (28 mg, 0.047 mmol) in dry methylene chloride (0.8 mL) containing triethylamine (20 μL). After stirring at -40°C for 2 hr, saturated NaHCO₃ solution (10 mL) was poured into the reaction mixture and this was extracted with ether (2 × 12 mL). Combined extracts were washed with brine twice, dried over MgSO₄-K₂CO₃, and concentrated *in vacuo*. The residual oil was separated by preparative TLC developing with 8% ethyl acetate-benzene containing 0.1% of triethylamine to isolate more polar component **5a** (5.3 mg, 17%) and less polar component **6a** (1.5 mg, 5%).

(5E,7S)-5,7-Dichloro-PGI₂ methyl ester (**5b**)

A solution of pyridine (0.14 mL) and hydrogen fluoride-pyridine (0.28 mL, Aldrich) in acetonitrile (2 mL) was prepared, and 0.4 mL of this solution was added to **5a** (18 mg, 0.027 mmol) at room temperature. After stirring at the same temperature for 1 hr, saturated NaHCO₃ solution (10 mL) was added and this was extracted

with ethyl acetate (2 × 12 mL). The combined extracts were washed with brine, dried over $\text{MgSO}_4 \cdot \text{K}_2\text{CO}_3$ and concentrated *in vacuo*. The residual oil was chromatographed on Florisil eluting with *n*-hexane-ethyl acetate (1 : 1) containing 0.1% of triethylamine to give **5b** (9 mg, 80%) as an oil; $[\alpha]_D^{22} + 98^\circ$ (c, 0.11, CHCl_3); IR (film) 3380, 1738, 1680, 1438, 1215, 1078, 970, 920, 655 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) δ 3.69(3H, s), 3.93(1H, q, $J=8$ Hz), 4.13(1H, q, $J=6$ Hz), 4.82(1H, s), 5.00(1H, m), 5.59(1H, dd, $J=8$, 16 Hz), 5.67(1H, dd, $J=6$, 16 Hz); MS m/e 436, 434(M^+), 400, 398, 382, 380, 363; Calc. for $\text{C}_{21}\text{H}_{35}\text{O}_5^{35}\text{Cl}$ 434.1625, Found 434.1648.

(5Z,7S)-5,7-Dichloro-PGI₂ methyl ester (6b)

The silyl-protecting groups of **6a** (20 mg, 0.030 mmol) were removed by the same method described above (as in the case of the desilylation of **5a**) to give 10 mg (76%) of **6b** as an oil; $[\alpha]_D^{22} + 107^\circ$ (c, 0.10, CHCl_3); IR (film) 3400, 1738, 1680, 1438, 1215, 1132, 972, 912, 652 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) δ 3.68(3H, s), 3.95(1H, q, $J=8$ Hz), 4.13(1H, q, $J=6$ Hz), 4.76(1H, s), 5.05(1H, m), 5.58(1H, dd, $J=8$, 16 Hz), 5.68(1H, dd, $J=6$, 16 Hz); MS m/e 436, 434(M^+), 400, 398, 380, 363, 345; Calc. for $\text{C}_{21}\text{H}_{31}\text{O}_5^{35}\text{Cl}$ (M-H ^{35}Cl) 398.1858, Found 398.1882.

Stepwise transformation of 2a into 7a and 8a via hemiketal 9

Pyridinium *p*-toluenesulfonate (20 mg) was added to a stirred solution of silyl-protected 5-chloro- δ^6 -PGI₁ **2a** (55 mg, 0.088 mmol) in 1 mL of moistured benzene. After stirring the mixture at room temperature overnight, saturated sodium bicarbonate solution was added and the mixture was extracted with ethyl acetate twice. The combined extracts were washed with brine, dried over $\text{MgSO}_4 \cdot \text{K}_2\text{CO}_3$ and concentrated *in vacuo* to give 47 mg of crude **9**. The thin layer chromatogram of **9** showed two spots due to the epimers at C-6; IR (CDCl_3) 3570, 3350, 1730, 1460, 1360, 1122, 1002, 972, 835 cm^{-1} ; NMR (CDCl_3) δ 0.7-1.0(21H), 3.67(3H, s), 3.7-4.1(3H, m), 4.5-4.8(1H, m), 5.43(2H, m); MS m/e 630, 628 (M-18), 592, 591, 589, 573, 571.

Anhydrous magnesium sulfate (1.2 g) was added to the crude hemiketal **9** (47 mg) in dry benzene (4 mL), and the mixture was stirred with reflux for 3 hr. After removing the magnesium sulfate with filtration, the filtrate was partitioned between ether and saturated sodium bicarbonate solution. The water layer was extracted with ether again, and the combined organic layer was washed with brine twice, dried over $\text{MgSO}_4 \cdot \text{K}_2\text{CO}_3$, and concentrated *in vacuo*. The residue (41 mg) was separated by preparative TLC developing with 8% ethyl acetate-*n*-hexane containing trace amount of triethylamine giving three components. **8a** (15 mg, 28%), **2a** (9 mg, 18%), **7a** (11 mg, 20%) in the order of polarity. The least polar **7a**; $[\alpha]_D^{23} + 11^\circ$ (c, 0.22, CHCl_3); IR (CHCl_3) 1736, 1696, 1462, 1260, 1122, 978, 838 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) δ 0.86(9H, s), 0.89(9H, s), 3.66(3H, s), 3.84(1H, q, $J=8$ Hz), 4.07(1H, m), 4.62(1H, m), 5.49(2H, m); $^{13}\text{C-NMR}$ (CDCl_3) δ 33.6 (C_2), 22.6(C_3), 32.9(C_4), 106.0(C_5), 153.2(C_6), 31.4(C_7), 44.8(C_8), 85.6(C_9), 41.7(C_{10}), 78.0(C_{11}), MS m/e 630, 628(M^+), 573, 571, 535, 439. Calc. for $\text{C}_{33}\text{H}_{61}\text{O}_5\text{Si}_2^{35}\text{Cl}$ 628.3743, Found 628.3734.

The most polar **8a**; $[\alpha]_D^{23} + 17^\circ$ (c, 0.24, CHCl_3); IR (CHCl_3) 1734, 1690, 1460, 1252, 1122, 975, 838 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) δ 0.86(9H, s), 0.89(9H, s), 3.66(3H, s), 3.85(1H, q, $J=7.6$ Hz), 4.07(1H, m), 4.70(1H, m), 5.48(2H, m); $^{13}\text{C-NMR}$ (CDCl_3) 33.4(C_2), 22.6(C_3), 32.7(C_4), 100.9(C_5), 151.2(C_6), 32.6(C_7), 45.5(C_8), 85.1(C_9), 41.6(C_{10}), 78.1(C_{11}), 54.6(C_{12}); MS m/e 630, 628(M^+), 573, 571, 535, 439. Calc. for $\text{C}_{33}\text{H}_{61}\text{O}_5\text{Si}_2^{35}\text{Cl}$ 628.3743, Found 628.3712.

Acid catalyzed isomerization of 2a into 7a and 8a

Trace amount of concentrated sulfuric acid was added by means of a thin glass capillary into a NMR sample tube containing a solution of **2a** (20 mg, 0.032 mmol) in chloroform-*d* (0.3 mL). As soon as the C-7 vinylic proton of the starting material was disappeared in its NMR spectrum, this mixture was poured into aqueous NaHCO_3 solution and extracted with methylene chloride twice. The combined extracts were washed with brine, dried over $\text{MgSO}_4 \cdot \text{K}_2\text{CO}_3$ and concentrated *in vacuo*. The residue was separated by preparative TLC developing with *n*-hexane-ethyl acetate (9 : 1) containing 0.5% of triethylamine giving less polar **7a** (2.0 mg, 10%) and more polar **8a** (2.0 mg, 10%).

(5E)-5-Chloro-PGI₂ methyl ester (7b) and its sodium salt solution (7c)

Tetrabutylammonium fluoride trihydrate (139 mg, 0.44 mmol) was added at room temperature to a stirred mixture of **7a** (18 mg, 0.028 mmol) and triethylamine (67 μL) in 2.5 mL of tetrahydrofuran. The reaction mixture was stirred at room temperature for 4 hr and concentrated *in vacuo*. Saturated NaHCO_3 solution (10 mL) was poured into the residue and this was extracted with ethyl acetate (2 × 12 mL). The combined extracts were washed with brine, dried over $\text{MgSO}_4 \cdot \text{K}_2\text{CO}_3$ and concentrated *in vacuo*. Chromatography of the crude product with Florisil eluted with *n*-hexane-ethyl acetate (3 : 1 ~ 1 : 1) containing 0.1% of triethylamine furnished 10.5 mg of **7b** (95%) as an oil; IR (CHCl_3) 3420, 1730, 1592, 1438, 1158, 1079, 972, 916 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) δ 2.16(1H, q, $J=8$ Hz; $\text{C}_{12}\text{-H}$), 2.325(2H, t, $J=7$ Hz; $\text{C}_2\text{-H}_2$), 2.37(1H, q, $J=8$ Hz; $\text{C}_3\text{-H}$), 2.38(1H, dt, $J=14$, 7 Hz; $\text{C}_4\text{-H}$), 2.48(1H, dt, $J=14$, 7 Hz; $\text{C}_8\text{-H}$), 2.48(1H, dt, $J=14$, 7 Hz; $\text{C}_{10a}\text{-H}$), 2.63(1H, d, $J=17$ Hz; $\text{C}_7a\text{-H}$), 2.70(1H, dd, $J=17$, 8 Hz; $\text{C}_9\text{-H}$), 3.68(3H, s), 3.90(1H, br; $\text{C}_{13}\text{-H}$), 4.10(1H, q, $J=6$ Hz, $\text{C}_{14}\text{-H}$), 4.66(1H, dt, $J=3$, 8 Hz; $\text{C}_5\text{-H}$), 5.53(1H, dd, $J=8$, 15 Hz; $\text{C}_{13}\text{-H}$), 5.64(1H, dd, $J=6$, 15 Hz; $\text{C}_{14}\text{-H}$); MS m/e 402, 400(M^+), 384, 382, 366, 365, 364, 347; Calc. for

$C_{21}H_{33}O_5^{35}Cl$ 400.2014, Found 400.1998.

To the solution of **7b** (8.7 mg, 0.022 mmol) in ethanol (1.3 ml) was added 1.3 ml of 0.1 M aqueous sodium hydroxide. The mixture was stirred at room temperature until the starting ester **7b** disappeared on TLC (6 h). This solution was used for pharmacological assay as a solution of **7c**.

(5Z)-5-Chloro-PGI₂ methyl ester (**8b**) and its sodium salt solution (**8c**)

The silyl-protecting groups of **8a** (22 mg, 0.035 mmol) were removed by the same method described above (as in the case of the desilylation of **7a**) to give 11 mg (80%) of **8b** as an oil; IR (CHCl₃) 3420, 1730, 1690, 1437, 1135, 1082, 975, 908 cm⁻¹; ¹H-NMR (CDCl₃) δ 2.17(1H, q, J=8 Hz; C₁₂-H), 2.27(2H, t, J=7 Hz; C₄-H₂), 2.33(2H, t, J=7 Hz; C₂-H₂), 2.44(1H, q, J=8 Hz; C₈-H), 2.52(1H, d, J=16 Hz, C_{7a}-H), 2.53(1H, dt, J=14, 8 Hz, C₁₀-H), 2.67(1H, dd, J=16, 8 Hz; C_{7b}-H), 3.67(3H, s), 3.91(1H, q, J=8 Hz; C₁₁-H), 4.10(1H, q, J=6 Hz; C₁₅-H), 4.74(1H, dt, J=3, 8 Hz; C₉-H), 5.52(1H, dd, J=8, 16 Hz; C₁₃-H), 5.64(1H, dd, J=6, 16 Hz; C₁₄-H); MS m/e 402, 400(M⁺), 384, 382, 366, 365, 364, 347; Calc. for C₂₁H₃₃O₅³⁵Cl 400.2014, Found 400.1980.

To the solution of **8b** (3.5 mg, 0.009 mmol) in ethanol (0.52 ml) was added 0.52 ml of 0.1 M aqueous sodium hydroxide. The mixture was stirred at room temperature until the starting ester **8b** disappeared on TLC (6 h). This solution was used for pharmacological assay as a solution of **8c**.

Fluorination of **3b** with silver fluoride—(7S)-7-fluoro-PGI₂ diacetate methyl ester (**16a**) and 5-fluoro-Δ⁸-PGI₂ diacetate methyl ester (**17a**)

Silver fluoride (610 mg, 4.8 mmol) was added at room temperature to a stirred solution of **3b** (253 mg, 0.48 mmol) in acetonitrile (12 mL) containing potassium carbonate (199 mg, 1.44 mmol) and the mixture was stirred at room temperature for 4 hr. Benzene (12 mL) was added and the mixture was filtered through Celite. After washing the Celite with benzene, combined filtrate was concentrated *in vacuo*. The resulting residue was purified by Florisil column chromatography to isolate two compounds; **16a** (90 mg, 40%) (eluting solvent: n-hexane-ethyl acetate, 9 : 1, 0.1% triethylamine) and **17a** (45 mg, 20%) (eluting solvent: n-hexane-ethyl acetate, 87.5 : 12.5, 0.1% triethyl amine). **16a**; IR (film) 1740, 1700, 1438, 1370, 1240, 970 cm⁻¹; ¹H-NMR (CDCl₃) δ 2.01(3H, s), 2.05(3H, s), 3.67(3H, s), 4.77(1H, t, J=7 Hz), 4.7-5.1(2H, m), 4.95(1H, d, J=5.5 Hz), 5.1-5.3(1H, m), 5.55(2H, m); ¹³C-NMR (CDCl₃) δ 33.5(C₂), 24.8(C₃), 24.8(C₄), 104.5(d, J=10.7 Hz; C₅), 153.3(d, J=15.3 Hz; C₆), 94.4(d, J=177.0 Hz; C₇), 52.1(d, J=22.9 Hz; C₈), 83.2(C₉), 37.9(C₁₀), 78.2(C₁₁), 48.3(d, J=6.1 Hz; C₁₂), MS m/e 468 (M⁺), 448, 408, 348, 328. Calc. for C₂₃H₃₃O₅F (M-ACOH) 408.2311, Found 408.2344. **17a**; IR (film) 1735, 1665, 1365, 1240, 970 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.99(3H, s), 2.04(3H, s), 3.14(1H, m), 4.6-5.3(5H, m), 5.51(2H, m); ¹³C-NMR (CDCl₃) δ 33.5(C₂), 20.3(d, J=3.7 Hz; C₃), 33.0 and 32.8(d, J=24.4 Hz; d, J=24 Hz, respectively: C₄), 87.54 and 87.47 (d, J=168 Hz; d, J=172 Hz, respectively: C₅), 154.5 and 154.4 (d, J=20.8 Hz; d, J=20 Hz, respectively: C₆), 101.2 and 101.7(d, J=6.1 Hz; d, J=7.3 Hz, respectively: C₇), 51.5, 54.5(C₈ or C₁₂), 84.7 and 84.9(C₉), 38.7(C₁₀), 77.9(C₁₁); MS m/e 468(M⁺), 448, 408, 348, 328.

Measurement of chemical stability in pH 2.35 buffer solution

A buffer solution was prepared by mixing 0.2 M-KCl (50 mL), 0.2M-HCl (4 mL) and ethanol (29 mL). The pH of this buffer solution was 2.35 at 23°C. Ethanol solutions (25 μL, 2 mg/ml) of chlorinated PGI₂ methyl esters **5b**, **6b**, **7b** and **8b** were added to the buffer solution (975 μL) at 23°C. The hydrolysis was monitored at the same temperature by following the peak area change of the compounds on high pressure liquid chromatography (HPLC) charts (Mobile phase, CH₃CN : H₂O = 1 : 1; Wave length, 235 nm for **5b** and **6b** and 210 nm for **7b** and **8b**). The half lives of PGI₂ methyl ester was measured in the same condition and was found to be less than one minute.

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